qDTY3.1, a major drought tolerant locus of APO promotes early flowering in the genetic back ground of a local cultivar improved white ponni

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Research Article

$\overline{qDTY3.1}$, a major drought tolerant locus of APO promotes early flowering in the genetic back ground of a local cultivar improved white ponni

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Abstract

Advancements in molecular genetics led to identification of genomic regions in rice associated with performance under drought conditions which enabled designing molecular breeding strategies for development of drought tolerant rice genotypes. However, physiological and molecular basis of most of these QTLs are not yet studied. In this study, a recombinant inbred population (RIL) developed between Improved White Ponni (a drought susceptible rice variety) and Apo (a drought tolerant cultivar) was genotyped using SSR markers linked to major drought tolerant QTLs. Evaluation of RILs for days to flowering and subsequent marker – trait association analysis revealed that qDTY3.1, a major drought tolerant loci was found to be associated with earliness with an R² value of 0.66. Further analysis revealed that qDTY3.1 was found to be co-localized or located near a major heading date locus Hd6 (30.4–35.6 Mb; RM3199–RM3329) on chromosome 3. Further experiments for developing back cross progenies harboring the two loci in different combinations are in progress to dissect out the interaction between the qDTY3.1 and Hd6.

Key words

Rice, Drought, SSR Marker, Validation

Introduction

Rice is the important food crop for more than half of the world's population. Abiotic stresses like drought and submergence severely affects rice production and productivity (Vikram et al., 2011). Among the various abiotic stresses, drought affects 23 million hectares of rainfed rice (Serraj et al., 2011) and thus causing severe threat to agricultural production (Pandey and Shukla 2015). When drought coincides with flowering and grain filling, it leads to drastic reduction in yield (Then et al., 2011). Developing drought tolerant rice varieties remains hampered due to complexity of tolerance mechanisms and lack of reliable phenotyping procedures. The identification of major QTLs for grain yield under droughtqDTY12.1 (Bernier et al., 2007), qDTY2.1 and qDTY3.1 (Venuprasad et al., 2009), and qDTY1.1 (Vikram et al., 2011) in rice using modern molecular biology tools has provided new opportunities to breeders to develop varieties tolerant against drought.

However, many of the QTLs reported to be associated with drought tolerance are 1) detected based on single marker analysis; 2) large in their size (>2 Mb interval) and 3) physiological and molecular basis are not yet understood and hence making it difficult to put them into practical breeding programs. Several attempts have been made at IRRI, Philippines to unravel physiological basis of these major effect QTLs and it has been reported that qDTY3.2 had major influence on days to flowering. In the present study, attempts were made through phenotyping and genotyping of a RIL population (F₃) derived between a popular long duration rice variety Improved White Ponni and a drought tolerant cultivar Apo to test the association between a major loci qDTY3.1 and days to flowering.

Materials and Methods

A set of recombinant inbred lines (F_3) derived between Improved White Ponni and Apo was used in this study. Improved White Ponni is a popular long duration variety possessing high yield and superior grain quality. Whereas Apo is a drought tolerant upland cultivar using which major QTLs viz., *qDTY1.1, qDTY1.2, qDTY 2.1, qDTY3.1, qDTY6.1* and *qDTY12.1* were mapped (Venu Prasad *et al.,* 2009; 2012a and 2012b). True F1s were identified and forwarded to F_2 generation using Improved White ponni as a recurrent and Apo as a donor parent. True F_{1s} were identified harboring *qDTY3.1* and forwarded to F_2 generation. In F3 generation, leaf samples were collected from 109 individual



progenies and used for extraction of genomic DNA through modified CTAB protocol as described by Murray and Thompson (1980). Isolated genomic DNA was quantified using Spectrophotometer (Nano Drop) and were diluted to final concentration of 50ng/ul, and stored under refrigerator condition at -40 ^oC.

A polymorphic SSR marker RM520 (30.9 Mb on chromosome 3) tightly linked to qDTY3.1 was used for genotyping of 109 F₃ progenies along with the parents. PCR reactions were carried out using DNA sample with the concentration of 50ng/ul, PCR buffer consist of 1.5mM MgCl₂, 0.4mM dNTPs, 10µM primers and 1.5U of Taq DNA polymerase were utilized. PCR reaction were performed in thermal cycler (DNA Engine, BioRad, USA) programmed as initial denaturation at 95°C for 5 min, 35 cycles of 95 °C for 30sec, 55 °C for 30 sec and 72 °C for 30 sec followed by final extension of one cycle at 72 °C for 10 min. PCR amplified products were subjected to gel electrophoresis using 3.5% agarose, stained with Ethidium Bromide and visualized with UV transilluminator. Observations on days to first flowering (emergence of spikelets of primary panicle outside the boot leaf sheath) and days to 50% flowering (when more than 50% of the panicles in a progeny started anthesis) were recorded.

Simple linear regression analysis (Zongo *et al.*, 2017) were carried out with the fallowing equation y = b0 + b1x + e (y- phenotypic value, b0 - population mean, b1x - function of molecular marker and <math>e = error) using marker and phenotypic value of a particular trait. The potential relationship can be judged by the phenotypic variance of \mathbb{R}^2 value (Boranayaka *et al.*, 2018).

Results and Discussion

Rice is one of the major cereals in Asia which remain at the top in both production and consumption (Hussain et al., 2014). Rice production is often hampered by several biotic/abiotic stresses and drought stands at the top among all the abiotic stresses in limiting rice yields especially under rainfed conditions (Prince et al., 2015). Rice crops depending on monsoon rains are frequently subjected to terminal drought and most of the existing varieties are highly sensitive to water deficit (Then et al., 2011). Developing early maturing rice varieties or varieties that can reduce their duration in response to drought will enable sustaining rice yields under drought. Several attempts have been made to unravel molecular genetic basis of drought tolerance traits in rice which led to identification of major loci governing performance under drought in a drought tolerant variety Apo (Venuprasad *et al.*, 2009), Nagina 22 (Vikram *et al.*, 2011) and Vandana (Solis *et al.*, 2018). However, only few of them including *qDTY1.1* and *qDTY12.1* have been dissected out at physiological and molecular level and put into breeding applications. Several other QTLs including *qDTY3.1* remain not fully understood at physiological and molecular levels.

In the present study, F_3 populations derived from the cross between Improved White ponni and Apo were subjected to precision genotyping and phenotyping with a view to study the association between *qDTY3.1* and days to flowering. A polymorphic marker RM520 located close to *qDTY3.1* (30.9 Mb on chromosome 3) was selected for genotyping (Fig.1; Table 1). A total of 109 F_3 progenies were genotyped using the marker RM520 which resulted in the identification of 32 progenies possessing homozygous IWP allele of RM520 and 34 progenies possessing homozygous Apo allele of RM520. Remaining 43 progenies were found to remain under heterozygous conditions.

Phenotyping of F₃ population for days to first flowering and days to 50% flowering revealed that progenies exhibited continuous variation for the traits. IWP was found to reach first flowering in 117 days and 50% flowering in 122 days (Table.2). Drought tolerant Apo registered first flowering in 108 days and 50% flowering in 112 days. F₃ progenies were found to reach first flowering ranging from 92 -113 days and 50% flowering between 96 - 119 days (Table.2). Progenies exhibited clear transgressive segregation which revealed the involvement of several loci in controlling the days to flowering (Fig 2). Marker vs trait analysis using simple linear regression method (Boranayaka et al., 2018) revealed that RM520 was found to possess significant association ($R^2 - 0.66$) with days to first flowering and days to fifty percent flowering (Table.3). Further, in silico analysis of qDTY3.1 revealed its close proximity or co-localization a major heading date locus, with Hd6 spanning between 30.4 Mb to 35.6 Mb on Chromosome 3 (Hori et al., 2015). Scanning of target regions using more number of SSR markers will allow us to group the lines based on recombination events between *qDTY3.1* and *Hd6*. Vikram et al. (2011) reported that *qDTY3.1* did not exhibit any association with days to flowering under normal condition in a population developed by involving Swarna and Apo. In this study, qDTY3.1 exhibited moderate association with days to flowering which may be due to the favorable



interaction of Apo with the recipient genome IWP. However, further confirmatory experiments are needed for dissecting out the precise role of qDTY3.1through development of backcross progenies of IWP harboring the QTL segment. This will also allow us to differentiate the effects of qDTY3.1 and Hd6 on promoting earliness in flowering and drought tolerance.

References

- Bernier, J., A. Kumar, V. Ramaiah, D. Spaner and G. Atlin. 2007. A large effect QTL for grain yield under reproductive stage drought stress in upland rice. *Crop Sci.J.*, **47**(2): 507-516.
- Boranayaka, M B., R. Lokesha, J.R. Diwan and R. Patil. 2018. Marker validation in F₂ population of rice (*Orza sativa* L.) for water and Nitrogen use efficiency. *Int. J. Curr. Microbiol. App. Sci.*, 7(1): 1275-1278.
- Hori, K., Y. Nonoue, N. Ono, T. Shibaya, K. Ebana, K. Matsubara, E.O. Tanaka, T. Tanabata, K. Sugimoto, F.T. Shiobara, J. Yonemaru, R. Mizobuchi, Y. Uga, A. Fukuda, T. Ueda, S. Yamamoto, U. Yamanouchi, T. Takai, T. Ikka, K. Kondo, T. Hoshino, E. Yamamoto, S. Adachi, H. Nagasaki, A. Shomura, T. Shimizu, I. Kono, S. Ito, T. Mizubayashi, N. Kitazawa, K. Nagata, T. Ando, S. Fukuka, T. Yamamoto and M. Yano. 2015. Genetic architecture of variation in heading date among Asian rice accessions. *BMC plant biology.*, 15:115.
- Hussain, S., T. Fujii, S. McGoey, M. Yamada, M. Ramzan and M. Akmal. 2014. Evaluation of different rice varieties for growth and yield characteristics. J. Anim. Plant Sci., 24(5): 1504-1510.
- Murray, M.G. and W.F. Thompson. 1980. Rapid isolation of high molecular weight plant DNA. *Nucleic Acid Res.*, **8(19):** 4321–4325.
- Pandey, V. and A. Shukla. 2015. Acclimation and tolerance strategies of rice under drought stress. *Rice Science.*, 22(4): 147-161.
- Prince, S J., R. Beena, S.M. Gomez, S. Senthivel and R.C. Babu. 2015. Mapping consistent rice (*Oryza* sativa L.) yield QTLs under drought stress in target rainfed environments. *Rice.*, 8:25.

- Serraj, R., K.L. McNally, I. Slamet-Loedin, A. Kohli, S.M. Haefele, G. Atlin and A. Kumar. 2011. Drought resistance improvement in rice: an integrated genetic and resource management strategy. *Plant Prod. Sci.*, **14**: 1-14.
- Solis, J., A. Gutierrez, V. Mangu, E. Sanchez, R. Bedre, S. Linscombe and N. Baisakh. 2018. Genetic mapping of quantitative trait loci for grain yield under drought in rice under controlled greenhouse conditions. *Front Chem.*, 5: 129.
- Then, R., J.L. Siangliw, A. Vanavichit, P. Kasemsap, S. Fukai and T. Toojinda. 2011. Effects of drought tolerant quantitative trait loci on flowering traits, panicle exsertion rate, spikelet sterility and grain yield of rice under rainfed lowland conditions. *Kasetsart J. (Nat. Sci.).*, **45:** 101-109.
- Venuprasad, R., M.E. Bool, L. Quiatchon and G.N. Atlin. 2012a. A QTL for rice grain yield in aerobic environments with large effects in three genetic backgrounds. *Theor. Appl. Genet.*, **124**: 323–332.
- Venuprasad, R., M.E. Bool, L. Quiatchon, M.T. Sta Cruz, M. Amante and G.N. Atlin. 2012b. A large effect QTL for rice grain yield under upland drought stress on chromosome 1. Mol. Breed., 30: 535– 547.
- Venuprasad, R., C.O. Dalid, M.D. Valle, D. Zhao, M. Espiritu, M.T. Sta Cruz, M. Amante, A. Kumar and G.N. Atlin. 2009. Identification and characterization of large effect quantitative trait loci for grain yield under lowland drought stress in rice using bulk segregant analysis. *Theor Appl Genet.*, **120(1)**: 177-90.
- Vikram, P., B.P.M. Swamy, S. Dixit, H.U. Ahmed, M.T.S. Cruz, A.K. Singh and A. Kumar. 2011. *qDTY1.1*, a major QTL for rice grain yield under reproductive-stage drought stress with a consistent effect in multiple elite genetic backgrounds. *BMC Genetics.*, **12:** 89.
- Zongo, A., P. Khera, M. Sawadago, Y. Shasidhar, M. Sriswathi, M.K. Vishvakarma, P. Sankara, B.R. Ntare, R.K. Varshney, M.K. Pandey and H. Desmae. 2017. SSR markers associated to early leaf spot disease resistance through selective genotyping and single marker analysis in groundnut (*Arachis hypogaea* L.). *Biotechnol Rep.*, **15**: 132-137.



QTL	Chr. No	Linked Marker	Marker interval	Physical position (Mb)	Reference
qDTY3.1	3	RM520	RM520 - RM16030	30.9	Venu Prasad et al.
			(30.9 – 32.5 Mb)		(2009)

Table 1. Description of target QTL (qDTY3.1) controlling yield under drought stress

Table 2. Performance of parents and F₃ lines (Improved white ponni x Apo) for days to heading traits

Genotypes	Allelic Pattern (<i>qDTY3</i> .1- RM520)	Days to first flowering (Days)	Days to fifty percent flowering (Days)	
IWP	AA	117	122	
APO	BB	108	112	
F ₃ progenies shown homozygous for IWP allele	AA	96-113	103-119	
F ₃ progenies shown homozygous for Apo allele	BB	92-111	96-118	

IWP - Improved White Ponni, AA - Homozygous for Improved White ponni allele, BB -Homozygous for Apo allele

Table 3. Marker trait association by regression analysis

Trait	Marker	Chromosome	Position	R² value
Days to first flowering	RM520	3	30.9	0.656
Days to fifty percent flowering	RM520	3	30.9	0.657





Fig.1. Foreground selection of F₃ population (Improved White ponni x Apo) using SSR primer (RM520) linked to the target QTL (*qDTY3.1*)



Fig.2. Frequency distribution of F_3 homozygous progenies for Improved white ponni and Apo allele for the trait days to fifty percent flowering



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